Environmental drivers of mitochondrial content and its role in feed efficiency and meat quality

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Session: Environmental factors can not be ignored in animal production

Overview

- Why tissue mitochondrial content?
 - <u>Feed efficiency</u> (low mitochondrial content, muscularity and high FE)
 - <u>Meat quality</u> (mitochondria and post-mortem metabolism?)
- Genetic and **environmental** drivers of high mitochondrial content
 - <u>Cold</u> ambient temperature (thermogenic tissues muscle and brown fat)
 - <u>Diet</u> high fat, ketone esters, curcumin, PQQ
 - <u>Activity</u> endurance exercise
- Illustrate with multiple production species, special focus on broilers



- Start a discussion about mitochondrial phenotypes in animal production and how they may be environmentally influenced in commercial settings:
 - Previous work with Cobb Vantress (broiler mitochondrial content variation)?
 - Previous work with USDA and University of Arkansas (molecular mechanisms of divergence in broiler FE, role for the mitochondrion)
 - New project proposal with Kemin and CAS (pig heat stress and molecular / mitochondrial mechanisms through which nutrition may provide mitigation)

Mitochondria and energy flow



A mitochondria is like a bicycle, converting one source of energy into another through a coupling mechanism.

Both the size of the bicycle and the coupling mechanism can be adapted to meet different metabolic goals.

What is a mitochondria?

- Complex, cellular organelle comprising ~1000 proteins
- The 'engine' of the cell
- Thought to be formerly a free-living bacteria that has formed a symbiotic relationship with the host cell of modern animals.



An animal cell: two genomes in cooperation

- Mitochondria retains some of its own genome, encoding 13 protein coding genes plus other machinery (DNA based molecular biotech can be used).
- Remaining (majority) mRNA encoding mitochondrial proteins are in the nuclear genome.
- The collective expression of the mRNA that encode the mitochondria is controlled by a combination of both environment and genetics



Mitochondria in action – brown versus white fat



Brown fat important for cold stressed neonatal mammals and has a likely role in feed efficiency

Why think about mitochondria?

- The 'engine' of the cell where the majority of feed energy is converted into biologically usable ATP (and some body heat).
- ATP ultimately pays for production, such as deposition of lean.
- Improvements in mitochondrial performance (food → ATP conversion) will improve whole animal feed efficiency.
- Improvements can be made by both environmental and genetic mechanisms.

ATP – the 5 cent coin of bioenergetics

- Dietary glucose, protein and fat can be seen \$100 notes
- If a biological process costs 10 cents, more efficient to pay with two 5 cent coins than breaking a big note and 'losing the change.'



Feed, digestion and energy conversion in the mitochondria



Where is the useful variation across populations of animals? Gut or **muscle**?

Mitochondrial adaptations, energy and heat



- Brown fat has a large bicycle (high energy flux) with no chain (no ATP, lots of heat).
- White fat has a small bicycle (low energy flux) with a chain (little heat, little ATP).
- Heart muscle has a large bicycle (high energy flux) with a chain (lots of ATP)

How do mitochondria convert energy?



Muscle mitochondrial content

- If the mitochondria is the engine of the cell, then mitochondrial content is engine size.
- Focus on muscle because it is 50% of the animals mass, a substantial contributor to whole animal metabolism, is composed of fibres with different mitochondrial contents and is the tissue we consume in animal production systems
- How might muscle mitochondrial content (engine size) influence production?
- What environmental factors drive mitochondrial content (engine size)

Feed efficient breeds tend to be built of low mitochondrial content type IIB muscle







'Corolla' chicken versus 'drag racing' hummingbird





Exceptional feed conversion Sedentary *Pectoralis* mitochondrial content **4%** Small engine 'Toyota Corolla' HibernationAthletic35%'Gas guzzling' drag racer

The transformed broiler – a ~50 year story



Genetics and environment in broiler production

- Genetic gain: breeding from genetically superior parents, now using DNA markers to inform the selection process
- Environmental change: nutrition and other management practices
- Collectively, improve the <u>efficiency of the flow of energy</u> into lean tissue deposition, primarily the enlarged breast muscle

Efficiency, flow, energy



Overview of recent broiler research

- Measuring broiler mitochondrial phenotypes (using DNA and RNA)
- Industrial (Cobb Vantress, HQ Arkansas, USA) and academic colleagues (University of Arkansas, USA)
- Both projects used Cobb Vantress broilers as the animal resource.

Cobb Vantress

- US multi-national company that produces broilers for people who produce broilers.
- Control the genetics for ~50% of the global supply of broilers.
- Genetic improvement with a particular emphasis on feed efficiency
- My role was to measure a cellular phenotype (mitochondrial content) that is the product of bird environment as well as bird genetics

Broiler mitochondrial content

- Screen birds (within a genetic line) for variation in muscle mitochondrial content. How much spread around the 4%?
- Develop the biotechnology to achieve this (qPCR from DNA).
- Connect any variation to 11 performance phenotypes (but not FE).

Broiler mitochondrial content



The assay can discriminate tissues with different metabolic demands.

There is a positive relationship (0.61; *P* < 0.0001) between breast and thigh content across 80 birds.

Mitochondrial content and performance traits

- Breast muscle mitochondrial content negatively correlated with:
 - Breast muscle yield (-0.27; *P* = 0.037)
 - Carcass yield (-0.26; *P* = 0.045)
 - Abdominal fat content (-0.31; *P* = 0.017)
- Thigh muscle mitochondrial content negatively correlated with:
 - Bow out leg defect (-0.30; *P* = 0.011)

Regulation of muscle mitochondrial content

Genetics

• species comparisons, domestication and industrial selection

Environment

- Exercise (endurance versus sprint exercise)
- Ambient temperature (heat versus cold)
- Diet (high fat, ketone esters, curcumin and PQQ)
- G and E integrated by a transcriptional regulator called PPARGC1A

Summary of Cobb Vantress research

- There appears to be systemic regulation of muscle mitochondrial content in chickens. A bird with a low value for thigh, will also have a low value for breast.
- A bird with low breast mitochondrial content tends to be more muscular with higher abdominal fat.
- A bird with low thigh mitochondrial content tends to be more likely to express the bow out leg pathology.

Mitochondrial content in broilers

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RESEARCH ARTICLE

Chicken muscle mitochondrial content appears co-ordinately regulated and is associated with performance phenotypes Antonio Reverter¹, Ron Okimoto², Robyn Sapp², Walter G. Bottie³, Rachel Hawken² and Nicholas J. Hudson^{4,*}

ABSTRACT

Mitochondrial content is a fundamental cellular bioenergetic phenotype. Previous work has hypothesised possible links between variation in muscle mitochondrial content and animal performance. However, no population screens have been performed in any production species. Here, we have designed a high throughput molecular approach to estimate mitochondrial content in commercial broilers. Technical validity was established using several approaches, including its performance in monoclonal DF-1 cells, cross-tissue comparisons in tissues with differing metabolic demands (white fat
breast muscle<drumstick muscle<heart muscle) and, as a negative control, a near absence of mtDNA amplification from whole blood. We screened breast muscle and thigh muscle in 80 birds individually phenotyped for 11 growth and development traits. Substantial individual variation (fivefold) was discovered in both breast and thigh muscle mitochondrial content. Interestingly, across birds we detected a very strong positive relationship between breast and thigh content (correlation coefficient 0.61; P<0.0001), consistent with coordinate regulatory control across the musculature. Further, breast muscle mitochondrial content is negatively correlated with breast muscle yield (-0.27; P=0.037), abdominal fat content (-0.31; P=0.017) and carcass vield (-0.26; P=0.045). Therefore, low breast muscle mitochondrial content is associated with more muscular birds possessing higher abdominal fat, the latter being in line with biomedical models of obesity. Finally, thigh mitochondrial content is

phenotypes. In the post-genomics era it has exploited single nucleotide polymorphism (SNP) genotyping to enable DNA marker-assisted selection practices (Lahav et al., 2006). The combination of the two practices has transformed domestic chickens over the last century to the modern animals with their precocious growth rates, dramatically increased muscle mass and markedly improved feed efficiencies (Paxton et al., 2010; Siegel, 2014).

The Company of Biologists

In principle, the DNA-based selection approach can mitigate the need to measure expensive phenotypes such as feed efficiency, or make predictions about individuals that do not directly express the phenotype, such as the contribution of male genetics to female reproductive characteristics. However, predicting phenotype from genotype is challenging even for industries that have an animal resource with a small effective population size and high levels of inbreeding. In chickens, the current accuracy of genomic prediction for a typical complex trait of moderate heritability is 54% (Lourenco et al., 2015). This is strong enough for implementation in a breeding strategy but there would be value in further improvements. One possible avenue is to develop biomarkers complementary to DNA sequence information. If these are practical and economical, they could be implemented in parallel to genetic testing and the two sources of information integrated for stronger predictions. Alternatively, any SNP subsequently found to be associated to the new biomarker could help refine the existing genomic predictions and therefore he delivered through the current DNA prediction

Q-CAS grant submitted February 2017

- Queensland (Uni Qld colleagues) and Chinese Academy of Science (Dr XiangFeng Kong) collaboration
- Kemin China (Dr. Bing Guo) are the industrial co-funder
- Explore the molecular basis of nutritional treatments (betaine and curcumin / essential plant oils) that may mitigate heat stress and improve production under sub-tropical conditions
- One part of the project is to develop a mitochondrial content assay for pigs, analogous to the industrial broiler research just described

University of Arkansas

• Prof. Walter Bottje, Centre of Excellence for Poultry Science



University of Arkansas and genome-wide gene expression

- Develop a deeper understanding of feed efficiency and its relationship to muscle structure and mitochondrial metabolism (Cobb broilers)
- Use genome-wide gene expression as the tool
- Observed patterns of gene expression are a complex consequence of environment and genetics

University of Arkansas and genome-wide gene expression

- Within a single genetic line, birds phenotyped for FE between 6 and 7 weeks, (1.5 fold difference).
- Breast muscle RNA extracted (n = 6 in each of two groups).
- Genome-wide mRNA quantified. This gives a global picture of ~10,000 gene expression values.
- Some of these genes encode proteins relating to muscle structure, mitochondrial function and metabolism.

Drowning in data

| 14001 11 10 120240 | | | 0.00002 | 0.04545 | 1.0021 | 0.10001 | 0.000 | 5.05020 | 5.05055 | 4.55011 | 5.11404 | 5.04515 | 0.00000 | 0.0 |
|---------------------|-------------|---|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|---------|------|
| 14502 A_73_120244 | FBX011 | Y | 12.3025 | 12.8356 | 12.4357 | 12.1377 | 12.3776 | 12.7111 | 12.4237 | 12.3494 | 12.3051 | 12.7244 | 12.4922 | 12.4 |
| 14503 A_73_120245 | LTB4R2 | Y | 6.06182 | 5.75777 | 5.99576 | 6.22621 | 5.65644 | 5.76563 | 5.68533 | 5.4969 | 5.97771 | 6.05889 | 6.14086 | 5.8 |
| 14504 A_73_120246 | MRPS24 | Y | 12.8914 | 13.2978 | 13.181 | 11.532 | 12.8201 | 12.723 | 13.3142 | 12.6336 | 13.0752 | 12.8598 | 13.3097 | 13.1 |
| 14505 A_73_120247 | RASAL2 | Y | 6.40008 | 6.04797 | 4.70939 | 5.5717 | 4.84951 | 5.08052 | 4.61057 | 4.3082 | 4.51624 | 4.84972 | 6.1868 | 5.5 |
| 14506 A_73_120249 | RAB7A | Y | 15.3736 | 16.1201 | 15.7261 | 15.3175 | 15.8505 | 16.1888 | 16.0098 | 15.6737 | 15.6727 | 16.1922 | 15.7201 | 15.6 |
| 14507 A_73_120250 | EIF2AK3 | Y | 6.69056 | 6.41431 | 6.03634 | 4.79695 | 5.63787 | 5.32609 | 4.70103 | 4.69475 | 4.91894 | 4.86378 | 7.03819 | 6.46 |
| 14508 A_73_120251 | NEDD4L | Y | 10.6408 | 8.82865 | 8.82306 | 9.3776 | 8.32224 | 9.06235 | 9.66553 | 9.11071 | 9.09552 | 9.59845 | 10.1862 | 9.10 |
| 14509 A_73_120253 | A_73_120253 | Y | 7.0448 | 6.98473 | 6.81664 | 6.00237 | 6.33256 | 6.82234 | 5.8353 | 5.64351 | 5.93596 | 6.6225 | 7.17678 | 7.12 |
| 14510 A_73_120254 | MSTN | Y | 8.72581 | 7.65877 | 8.63289 | 4.93011 | 7.79136 | 8.93952 | 8.98846 | 6.96958 | 7.77954 | 7.00617 | 9.34228 | 8.65 |
| 14511 A_73_120255 | C14H8ORF70 | Y | 7.50045 | 6.87585 | 7.0741 | 3.64667 | 4.34447 | 4.72573 | 4.7688 | 4.15779 | 4.47055 | 4.46076 | 6.97353 | 7.15 |
| 14512 A_73_120257 | GIMAP7 | Y | 2.53541 | 5.41354 | 4.88169 | 4.49988 | 6.98193 | 7.31226 | 7.57265 | 7.05273 | 7.49784 | 5.99213 | 2.99342 | 3.87 |
| 14513 A_73_120258 | ALDH7A1 | Y | 10.4526 | 9.71446 | 8.7367 | 7.75682 | 8.95604 | 9.97182 | 9.57148 | 9.4855 | 9.38248 | 8.95401 | 9.97283 | 9.20 |
| " 14514 A_73_120259 | ADAMTS10 | Y | 6.7161 | 6.10551 | 6.40848 | 4.97768 | 4.65453 | 4.31532 | 3.57451 | 3.92325 | 4.31422 | 5.3358 | 6.48577 | 6.2 |
| 14515 A_73_120261 | HSD17B7 | Y | 12.0477 | 11.6968 | 10.9042 | 10.0601 | 9.76593 | 9.7111 | 10.2038 | 9.87441 | 10.2706 | 10.4067 | 11.8753 | 11 |
| 14516 A_73_120262 | MKI67IP | Y | 10.4829 | 10.5049 | 9.43027 | 11.204 | 9.22997 | 9.02327 | 9.68939 | 8.7211 | 9.34814 | 9.54323 | 10.875 | 10.1 |
| 14517 A_73_120263 | A_73_120263 | Y | 9.45541 | 7.86952 | 7.24895 | 6.18001 | 7.27421 | 6.98965 | 7.59376 | 7.37168 | 7.48281 | 7.82077 | 6.68413 | 5.5 |
| 14518 A_73_120264 | KLHL21 | Y | 5.49553 | 5.61282 | 4.67782 | 4.36253 | 3.85963 | 3.38521 | 3.3125 | 2.94191 | 3.69872 | 3.72848 | 5.88035 | 5.24 |
| | | | | | | | | | | | | | | |



University of Arkansas and genome-wide gene expression I





University of Arkansas and genome-wide gene expression II





| Location | Gene ID | High FE - LowFE |
|----------------|----------------|-----------------|
| thick filament | MYH15 MYL2 | -2.16 -1.08 |
| | МҮОМЗ | -1.03 |
| thin filament | ТРМЗ | -1.75 -1.42 |
| z-disk | MYOZ2 CSRP3 | -1.34 -1.39 |
| | | |

A 3d muscle model can be used to asses the physical structure of muscle

Fibre composition has changed, less slow, red fibres in the more efficient birds

University of Arkansas and genome-wide gene expression III



Panel of 40 genes expressed in breast muscle can discriminate the 12 birds into treatment group i.e. high versus low efficient

More efficient birds express less slow, red muscle (e.g. MYL2, MB, TNNI1, MYOZ2, TPM3, MYH15) and have altered mitochondrial metabolism (e.g. CKMT1A).

Differential network analysis



Hudson et al (2009). *PLoS Computational Biology*. 5(5):e100382

University of Arkansas and genome-wide gene expression IV



Progesterone signaling

- Progesterone added to some Hormone Growth Promotant mixes in cattle, elevate FE by 20%.
- Progesterone changes physiology and fibre composition phenotype of muscle in pregnant women.
- Progesterone receptor co-localises to the mitochondria in birds.

Progesterone receptor immuno-staining



Summary of University of Arkansas research

- Panel of 40 genes expressed in muscle that can discriminate the two groups of birds differing in FE
- More efficient birds have paler muscle, a bias towards type II sprint fibres and altered mitochondrial function.
- Progesterone signalling is predicted to be a driver of the difference observed in muscle gene expression.
- Some feeds are high in progesterone-like compounds (yams)

Molecular mechanisms of feed efficiency in broilers

Bottje et al. BMC Systems Biology (2017) 11:29 DOI 10.1186/s12918-017-0396-2

BMC Systems Biology

RESEARCH ARTICLE

Open Access



Progesterone signalling in broiler skeletal muscle is associated with divergent feed efficiency

Walter Bottje¹, Byung-Whi Kong¹, Antonio Reverter², Ashley J. Waardenberg^{2,3}, Kentu Lassiter¹ and Nicholas J. Hudson^{4*}

Abstract

Background: We contrast the *pectoralis* muscle transcriptomes of broilers selected from within a single genetic line expressing divergent feed efficiency (FE) in an effort to improve our understanding of the mechanistic basis of FE.

Results: Application of a virtual muscle model to gene expression data pointed to a coordinated reduction in slow twitch muscle isoforms of the contractile apparatus (*MYH15*, *TPM3*, *MYO22*, *TNN11*, *MYL2*, *MYOM3*, *CSRP3*, *TNNT2*), consistent with diminishment in associated slow machinery (myoglobin and phospholamban) in the high FE animals. These data are in line with the repeated transition from red slow to white fast muscle fibres observed in agricultural species selected on mass and FE. Surprisingly, we found that the expression of 699 genes encoding the broiler mitoproteome is modestly–but significantly–biased towards the high FE group, suggesting a slightly elevated mitochondrial content. This is contrary to expectation based on the slow muscle isoform data and theoretical physiological capacity arguments. Reassuringly, the extreme 40 most DE genes can successfully cluster the 12 individuals into the appropriate FE treatment group. Functional groups contained in this DE gene list include metabolic proteins (including opposing patterns of *CA3* and *CA4*), mitochondrial network method (Regulatory Impact Factors) whose aim is to use patterns of differential co-expression to detect regulatory molecules transcriptionally rewired between the groups. This analysis clearly points to alterations in progesterone

Q-CAS grant submitted February 2017

- Queensland (Uni Qld colleagues) and Chinese Academy of Science (Dr XiangFeng Kong) collaboration
- Kemin China (Dr. Bing Guo) are the industrial co-funder
- Explore the molecular basis of nutritional treatments (betaine and curcumin / essential plant oils) that may mitigate heat stress and improve production under sub-tropical conditions
- One part of the project is to use genome-wide gene expression, analogous to the academic broiler research just described

Overall summary of broiler research

- There is substantial variation in muscle mitochondrial content in broilers and it associates with performance phenotypes including muscle and fat development.
- Patterns of gene expression indicate more efficient broilers have altered muscle structure and mitochondrial metabolism.

White striping and wooden breast



- Broiler breast muscle has an extreme muscle structure and metabolism
- Lactic acidosis has been observed in live birds predisposed to these conditions, implying inadequate aerobic metabolism
- **Hypothesis**: Is this a consequence of too low a mitochondrial content and capillarity?

Post mortem pH decline and meat quality



Conventional model relates to stored carbohydrate available for muscle glycolysis and subsequent anaerobic metabolism \rightarrow pH decline rate and ultimate pH

Given post-slaughter muscle is an enforced anaerobic systems is their a role for the (aerobic) mitochondria?

Mitochondrial 'treason' during anoxia

- In the absence of oxygen, mitochondria become net consumers rather than producers of ATP
- May hasten anaerobically produced ATP thereby influencing the pH decline curve





Conclusions

- Mitochondrial content varies across individual broilers and is associated with commercial performance traits
- Patterns of gene expression indicate high FE broilers have paler muscle and altered mitochondrial metabolism
- Mitochondrial phenotypes are responsive to environmental cues (ambient temperature, exercise and diet).
- Should the Q-CAS grant (co-funded by Kemin) be funded we have the opportunity to explore some of these ideas in the context of nutritional manipulation of pig production under heat stress

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